endoTest
Endoscope Microbiological Sample Kit

Instructions for Use New Endotest V2 with Hand Pump

A. Confirm insertion tube size corresponds with sample collection set.
B. Confirm accessory set applies to the scope to undergo the microbiological scope test.

Materials Required

1. Main collection set vessel ET20XX
2. Collection accessories set ETAKXXM
3. Bottle of sterile water
4. Sterile gloves
5. Clamp block/hand pump
6. Alcohol Towels - recommended

Set Up Instructions

All surfaces, clamp and hand pump should be wiped over between each test using in house disinfectant wipes

Main Collection Test Unit

1. Place clamp if available on working surface.
2. Obtain blister pack containing 120ml collection vessel set, note batch number located on Tyvek lid.
3. Removing the Tyvek lid to open the pack, obtain the supplied plastic sheet, unwrap, and use as your work area.
4. Scope clamp - ensure the plastic sheet is over clamp area where the scope is to be seated. Pull down over the upstand piercing the field to allow the sheet to sit correctly. Re wipe upstand with disinfectant wipes
5. Obtain your scope, lay scope flat on the draped sheet surface in looped formation with light guide post end facing you, place the scope handle in clamp and secure with a white strap, your scope LGT should be seated around the outside of the upstand SS bar.
6. Remove 300mm tube and 120ml vessel from the pack, check silicon seals are seated correctly and have not moved during transit.
7. Place the 120ml vessel onto the upstand located on the scope clamp facing the draped sheet area
8. Place the 300mm tube onto the tube section seated on the side of collection vessel.
9. Obtain your blue pump, attach the other end of the 300mm tube to the pump tube section.

Connection of scope to Collection Pot and Accessory Kit

For large scopes, older scopes or scopes that feel like they have resistance it is recommended to wipe around the IT bending rubber with an alcohol moist towel allowing lubrication. This will allow the scope to enter the silicon fitting easier.

1. Handle insertion tube approx. 1-2 inches from tip depending on scope (wipe with alcohol if required). Immediately after alcohol insert scope tip into the collection vessels silicon insert by gently pushing the tip of scope until the tip protrudes approx. 10mm into the collection vessel, **must not go past the entry of the specimen jar section.**
    - Small diameter scopes – lock angulation if possible. This will help to avoid angulation moving and make it easier while inserting
2. Once inserted check tip is facing down into the specimen jar. If need angulate the scope tip down this will allow the brush to enter the pot to reach the sample with ease during the brushing sequence.
3. Obtain bottle of sterile water - place bottle of water in centre of sterile field close to the scope handle

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Accessory Kit

4. Obtain the blister accessory pack.
5. Open accessory kit, obtain brush(s) and place aside for brushing sequence. "Sterile" do not open until ready for brushing sequence.
6. Remove manifold tube set being careful not to contaminate the bottle line that will enter the sterile water.
   Pick up the set using the manifold section. Immediately place bottle line into the bottle, and secure manifold to bottle using the bottle clip.
7. Proceed to attach the rest of manifold lines to the scope ports as per the accessory set instructions.
8. Once all connected, attach any channel blockers supplied in the set.

Method for gathering sample.

After connecting the relevant accessory set, ensure all connections and caps/plugs are secure.

1. Obtain your hand pump with the 300mm tube connected
2. **Refer to appropriate scope step by step colour coded collection method chart.**
   - Proceed to irrigate/flush all channels individually by selecting the appropriate channel clamp **as per colour coded chart.** Start with brushing channels being suction and biopsy (Yellow and White clamp lines), then proceed to small channels Red and Blue.

3. Using the hand pump ½ pump as instructed until fluid is witnessed entering the pot then close the channel clamp after approximately 10ml has entered the pot. (measurements on side of specimen jar**). Be mindful of how much fluid is being collected during the first collection step. Pump slowly - vessel can fill up quickly.**

Brushing Channels – This is your aseptic sterile part of the test

1. Remove the suction cylinder port cap as well as the biopsy port connection (white line) place on the plastic field to allow for brushing of the large channels as per the GENCA Guidelines.
2. Open brush without touching the brush and place safely on the plastic field ready for use.
3. Follow the aseptic method for brushing, hand hygiene and using sterile gloves.
4. Brush all brush able channels.
   a. 1st channel insert 90 degrees being the suction channel ensuring you site the tip of the brush in the silicon tube at the end of the yellow line connected at the suction port.
   b. 2nd channel, again the suction cylinder but insert @ 45 degrees and brush to approx. 200mm to go past the biopsy port. Remove brush.
   c. 3rd insert into biopsy port and brush until you see the tip of brush in the collection pot fluid sample – wash brush in fluid in collection pot by applying agitation.
   d. Remove brush and discard.
5. Replace the cylinder cap and biopsy connection, then proceed to flush and collect the brushing samples from the suction (yellow) and biopsy (white) channels.

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Completion of test

1. Using the hand pump collect brushing samples from brush able channels.
    a. Start with the yellow then white
    b. Small diameter you can open both yellow and white together if desired.

As we are completing the test, **you need to remove the bottle line from the water bottle as soon as you witness the fluid sample entering the collection pot.**

2. To collect sample, use hand pump till you here air being sucked through the channels. Close yellow
3. Open white and collect again until no fluid enters the collection vessel.
    Do large channels, you can do smaller blue/red channels if you feel the need and can do them together if you like.

Removing scope and sealing sample

1. **Carefully remove complete the silicon insert with the scope tip still attached**, place on working field.
2. Remove 300mm silicon tube from side of collection pot
3. Slide collection vessel off the clamp stands careful not to tip.

4. Sit on a flat surface. With hand flat on top push and wiggle down to seat the 120ml jar with fluid sample into up to join the lid, once seated twist to lock lid onto jar.

5. To remove the 120ml sample jar from the V2 collection vessel, wrap fingers around the silicon base ring and pull 120ml sample jar down with a swivel method till it has come away from V2 jar. Discard used seal.
   You can utilise the alcohol wipes by wiping the outside jar to lubricate so the silicon slides off easier

6. Check that the yellow lid is sealed tight.

7. Fill out test ID label (supplied) or your own department printed labels, and place on 120ml jar.
   Place collection vessel and appropriate pathology paperwork into sealed specimen bag and prepare to ship to lab for testing.

8. Disconnect all accessory connections caps/plugs from scope ports and discard all non-required parts.

9. **Remove silicon insert from tip of scope. (If, required use alcohol towel around IT before removing for lubrication. Wipe from end of silicone fitting down to tip while removing) discard.**